

Package ‘scCAN’

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Type Package

Title Single-Cell Clustering using Autoencoder and Network Fusion

Version 1.0.1

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Description A single-cell Clustering method using 'Autoencoder' and Network fusion ('sc-CAN') for segregating the cells from the high-dimensional 'scRNA-Seq' data. The software automatically determines the optimal number of clusters and then partitions the cells in a way such that the results are robust to noise and dropouts. 'sc-CAN' is fast and it supports Windows, Linux, and Mac OS.

License LGPL

Encoding UTF-8

LazyData true

LazyDataCompression xz

Depends R (>= 3.4), scDHA, FNN

Imports purrr, stats, markdown

RoxygenNote 7.1.1

Suggests knitr

VignetteBuilder knitr

NeedsCompilation no

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Repository CRAN

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adjustedRandIndex	<i>adjustedRandIndex</i>
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Description

The function to calculate adjusted Rand index value with the inputs of true clusters and predicted clusters

Usage

```
adjustedRandIndex(x, y)
```

Arguments

x	A vector that contain predicted cluster assignment.
y	A vector that contain true cluster assignment.

scCAN	<i>scCAN</i>
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Description

This is the main function to perform sc-RNA seq data clustering clustering. scCAN is fully unsupervised scRNA-seq clustering framework that uses deep neural network and network fusion-based clustering algorithm. First, scCAN applies a non-negative autoencoder to filter scRNA-seq data. Second, the filtered data is passed to stacked Bayesian autoencoder to get multiple low-dimensional representations of input data. Subsequently, scCAN converts these compressed data into networks and unify those networks to a single graph. Then, scCAN uses a spectral clustering algorithm to obtain final clusters assignment.

Usage

```
scCAN(
  data,
  sparse = FALSE,
  n.neighbors = 30,
  alpha = 0.5,
  n.iters = 10,
  ncores = 10,
  r.seed = 1
)
```

Arguments

<code>data</code>	Gene expression matrix, with rows represent samples and columns represent genes.
<code>sparse</code>	Boolean variable indicating whether data is a sparse matrix. The input must be a non negative sparse matrix.
<code>n.neighbors</code>	Number of neighboring cells that are used to calculate the edge's weight. The number of neighbors are set <code>n.neighbors = 30</code> by default.
<code>alpha</code>	A hyper-parameter that is used to calculate the network kernel. The value is set to <code>alpha = 0.5</code> by default.
<code>n.iters</code>	A hyper-parameter to set the number of network fusion iterations. It is set to <code>n.iters = 10</code> by default.
<code>ncores</code>	Number of processor cores to use.
<code>r.seed</code>	A parameter to set a seed for reproducibility. This values is set to <code>r.seed = 1</code> by default.

Value

List with the following keys:

- `cluster` - A numeric vector containing cluster assignment for each sample.
- `k` - The optimal number of cluster.

References

1. Duc Tran, Hung Nguyen, Bang Tran, Carlo La Vecchia, Hung N. Luu, Tin Nguyen (2021). Fast and precise single-cell data analysis using a hierarchical autoencoder. Nature Communications, 12, 1029. doi: 10.1038/s41467-021-21312-2

Examples

```
# Load the package and the example data (SCE dataset)
library(scCAN)
#Load example data
data("SCE")

#Get data matrix and label
data <- t(SCE$data); label <- as.character(SCE$cell_type1)

#Generate clustering result, the input matrix has rows as samples and columns as genes
result <- scCAN(data, r.seed = 1)

#Get the clustering result
cluster <- result$cluster

#Calculate adjusted Rand Index
ari <- round(scCAN::adjustedRandIndex(cluster,label), 2)
print(paste0("ARI = ", ari))
```

SCE

SCE

Description

SCE dataset includes scRNA-seq data and cell type information.

Usage

SCE

Format

An object of class `list` of length 2.

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